

# BIOINFORMATICS SESSION 12. PRACTICE

2023-11-20

Finding genes: going ashore at CpG islands

# Basic Shell Commands

```
$ cd [User_Folder]  
$ mkdir session12  
$ cd session12
```

# Basic Shell Commands

```
$ time
```

```
[biguser@R440 session11]$ time  
  
real    0m0.000s  
user    0m0.000s  
sys     0m0.000s
```

```
$ time python [script]
```

```
[biguser@R440 session12]$ time python cpg.py short.fa > $(date '+%Y-%m-%d').log  
  
real    0m0.194s  
user    0m0.110s  
sys     0m0.095s
```

# Basic Shell Commands

```
$ date # Current time and date
```

```
[biguser@R440 session11]$ date  
Mon Nov 13 16:30:33 KST 2023
```

```
$ date '+%F %r' # %F : YYYY-MM-DD  
# %r : 12 hour format
```

```
[biguser@R440 session11]$ date '+%F %r'  
2023-11-13 04:31:27 PM
```

```
$ date '+%Y-%m-%d' # YYYY-MM-DD (same as %F)
```

```
[biguser@R440 session11]$ date '+%Y-%m-%d'  
2023-11-13
```

# Basic Shell Commands

```
$ time python [script] > $(date '+%Y-%m-%d').log
```

```
[biguser@R440 session12]$ time python cpg.py short.fa > $(date '+%Y-%m-%d').log  
real    0m0.194s  
user    0m0.110s  
sys     0m0.095s
```

# Code 15.1

## cpg.py

```
1 import sys
2 import re
3
4 win = 500
5 step = 10
6 seq = ''
7
8 input_file = sys.argv[1]
9 for line in open(input_file): # a shorter sequence
10     if not re.search('>', line):
11         line = line.rstrip()
12         seq = seq + line
13
14 print('pos\tcpg\tcg_ratio\tcg_obs_exp')
15
16 for i in range(0, len(seq) - win+1, step):
17
18     testseq = seq[i:i + win]
19     c = float(testseq.count('C'))
20     g = float(testseq.count('G'))
21     cg = float(testseq.count('CG'))
22     cg_ratio = (c + g) * 100 / len(testseq)
23     cg_obs_exp = cg * len(testseq) / (c * g)
24     pos = i + win / 2
25     if cg_ratio >= 55 and cg_obs_exp >= 0.65:
26         print(str(pos)+ '\t'+ "1"+ '\t'+ str(cg_ratio)+'\t'+ str(cg_obs_exp))
27     else:
28         print(str(pos)+ '\t'+ "0"+ '\t'+ str(cg_ratio)+'\t'+ str(cg_obs_exp))
```

$$\frac{\frac{cg}{\text{len}(\text{testseq})}}{\frac{c}{\text{len}(\text{testseq})} \times \frac{g}{\text{len}(\text{testseq})}}$$

CG ratio      Observed ratio

C ratio      G ratio      Expected ratio

# Code 15.1

## cpg.py

```
$ cp /home/biguser/tutor/session12/short.fa .  
$ python cpg.py short.fa > cpg.short.out  
$ less cpg.short.out
```

pos	cpg	cq_ratio	cg_obs_exp
250.0	0	36.2	0.13354700854700854
260.0	0	36.4	0.13149243918474687
270.0	0	36.8	0.12756729174639622
280.0	0	37.0	0.18972931950417404
290.0	0	37.4	0.18315018315018314
300.0	0	36.6	0.1918158567774936
310.0	0	36.8	0.1926040061633282
320.0	0	37.0	0.19230769230769232
330.0	0	37.2	0.19211065573770492
340.0	0	37.2	0.25614754098360654
350.0	0	37.6	0.24838549428713363
360.0	0	38.4	0.2945681630729351
370.0	0	38.2	0.29698265621287717
380.0	0	38.2	0.29890004782400764
390.0	0	38.2	0.29698265621287717
400.0	0	38.6	0.29036004645760743
410.0	0	38.0	0.2976190476190476
420.0	0	37.8	0.30193236714975846
430.0	0	37.6	0.3044696139325295
440.0	0	37.2	0.30967422271770095
450.0	0	37.0	0.312343828085957
460.0	0	36.4	0.3244646333549643
470.0	0	37.4	0.3052503052503053
480.0	0	37.2	0.30458089668615984
490.0	0	37.8	0.29677113010446343
500.0	0	38.4	0.2863032524049473

4350.0	0	60.4	0.5649717514124294
4360.0	0	61.0	0.5763944311430345
4370.0	0	61.6	0.6076388888888888
4380.0	1	61.8	0.6538796861377506
4390.0	1	63.0	0.6652806652806653
4400.0	1	63.2	0.6984674801758495
4410.0	1	62.8	0.7061853528849749
4420.0	1	63.6	0.7285974499089253
4430.0	1	63.6	0.7315586262954684
4440.0	1	63.2	0.719010641357492
4450.0	1	63.6	0.7045088566827697
4460.0	1	64.4	0.7260596546310832
4470.0	1	65.0	0.7108276339045569
4480.0	1	65.4	0.7000681147354878
4490.0	1	66.4	0.7120166502355132
4500.0	1	66.8	0.7017165065313613
4510.0	1	67.0	0.6983240223463687
4520.0	1	66.4	0.7107709130672498
4530.0	1	66.0	0.7373271889400922
4540.0	1	65.8	0.7421150278293135
4550.0	1	66.0	0.7736516357206012
4560.0	1	65.4	0.7876969242310577
4570.0	1	65.4	0.7882291119285338
4580.0	1	65.2	0.773001508295626
4590.0	1	66.2	0.8048584180873637
4600.0	1	66.8	0.8257638315441783

# Visualization of CpG landscape with R

```
$ cp /home/biguser/tutor/Week12/cpg_short.r .  
$ vi cpg_short.r
```

```
# plot the results of cpg island prediction  
  
# define some colours  
rgb <- c("#000000", "#E69F00", "#56B4E9", "#009E73",  
         "#F0E442", "#0072B2", "#D55E00", "#CC79A7")  
  
# read file being output from Perl script  
data <- read.table("cpg_short.out", sep = "\t", header = TRUE)  
  
# Specify the number of lines of margin to the four sides  
# of the plot. In this case we want to make room for text  
# at the right axis.  
  
pdf("cpg_plot.pdf")  
par(mar = c(5, 4, 4, 5) + 0.1)  
  
# make first plot, which is empty  
plot(data$pos, data$cg_obs_exp, type = "n", xaxt = "n",  
      yaxt = "n", xlab = "", ylab = "")  
  
# plot lines to indicate where CpG islands are predicted  
for (i in 1:length(data$pos)) {  
  if (data$cpg[i] == 1) {  
    lines(c(data$pos[i], data$pos[i]), c(0, 1), col = rgb[2])  
  }  
}
```

⋮



# Visualization of CPG landscape with R

```
# before another plot, prevent R from clearing
# the graphics device
par(new = TRUE)

# make second plot with the cg_obs_exp data
plot(data$pos, data$cg_obs_exp, type = "l", main = "CpG island prediction",
      xlab = "Position", ylab = "CpG obs/exp", col = rgb[7])

# print legend
legend(7000, 0.9, c("CG ratio", "CpG obs/exp"), col = c(rgb[6],
  rgb[7]), lwd = 2)

par(new = TRUE)

# make third plot
plot(data$pos, data$cg_ratio, type = "l", xaxt = "n",
      yaxt = "n", xlab = "", ylab = "", col = rgb[6])

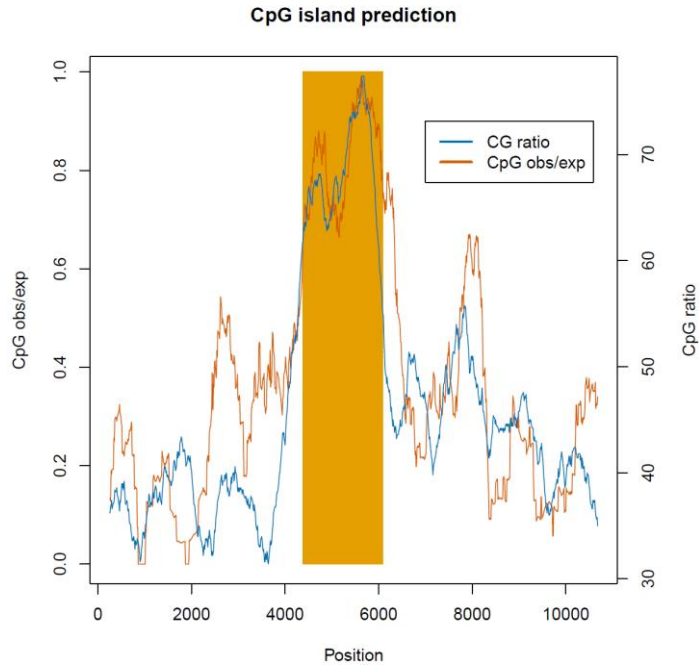
# print ticks for the 2nd y axis
axis(4)

# print text to 2nd y axis
mtext("CpG ratio", side = 4, line = 3)
dev.off()
```

# Visualization of CpG landscape with R

```
$ Rscript cpg_short.r
```

cpg\_plot.pdf



# Visualization of CPG landscape with R

```
$ cp /home/biguser/tutor/session12/chr4_region.fa .
$ python cpg.py chr4_region.fa > cpg_chr4.out
$ cp /home/biguser/tutor/session12/cpg_chr4.r .
$ cp /home/biguser/tutor/session12/chr4_annotation.txt .
$ less -S chr4_annotation.txt
```

chr	name	category	beg	end	z	strand	none	gene_id			gene_id	transcript_id
chr4	hg19_knownGene	exon	72053003	72053118	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72102293	72102366	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72120937	72121116	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72205087	72205222	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72215629	72215789	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72222725	72222904	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72263294	72263370	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72306333	72306490	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72313363	72313450	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72316106	72316260	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72316905	72317018	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72319212	72319386	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72332161	72332294	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72338416	72338687	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72352665	72352735	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72363218	72363409	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72397779	72397892	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72399944	72400105	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72412067	72412245	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72413365	72413437	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";
chr4	hg19_knownGene	exon	72420857	72420925	0.000000	+	.	.	.	.	gene_id "uc003hfy.2";	transcript_id "uc003hfy.2";

# Visualization of CpG landscape with R

## Step 1: CpG island plot

```
$ vi cpG_chr4.r
# plot the results of CpG island prediction

#define some rgb colours
rgb <- c("#000000", "#E69F00", "#56B4E9", "#009E73",
         "#F0E442", "#0072B2", "#D55E00", "#CC79A7")

# read data which is output of Perl script
data <- read.table("cpG_chr4.out", sep = "\t", header = TRUE)

# make an empty plot
plot(0, type = "n", xlim = c(72009075, 72009075 +
                             1520980), ylim = c(1.6, 2.3), xlab = "Position", ylab = "",
     yaxt = "n", main = "CpG island prediction")

# plot the predicted CpG islands
for (i in 1:length(data$pos)) {
  if (data$cpg[i] == 1) {
    # convert the position numbers to chromosomal positions
    data$pos[i] <- data$pos[i] + 72009075
    #print (data$pos[i])
    lines(c(data$pos[i], data$pos[i]), c(1.7, 1.8), col = rgb[7])
  }
}
⋮
```

# Visualization of CPG landscape with R

## Step 2: Gene structure plot

```
# Read file with chr4:72,009,075-73,530,562 region
# annotation. This information was obtained with
# the Table Browser of the UCSC browser

annot <- read.table("chr4_annotation.txt", sep = "\t",
                    header = TRUE)

color <- 1
prevname <- ""
lines <- length(annot$chr) # number of lines in the annotation file

for (i in (1:lines)) {

  if (annot$category[i] == "exon") {
    # if we consider an exon
    # if it a different gene as compared to the previous line,
    # change the colour
    if (annot$gene_id[i] != prevname) {
      color <- color + 1
    }
    prevname <- annot$gene_id[i]

    # draw rectangles for the exons
    rect(annot$beg[i], 1.9, annot$end[i], 2.1, border = rgb[color],
        col = rgb[color])
  }
}
```

`rect()`: plot region 안에 네모 모양(상자)을 그리는 함수.

```
function (xleft, ybottom, xright, ytop, density = NULL, angle = 45,
         col = NA, border = NULL, lty = par("lty"), lwd = par("lwd"), ...)
```

<b>xleft</b>	: 사각형의 왼쪽 x좌표.	<b>lty</b>	: 선의 종류 (테두리 및 내부 빗금)
<b>ybottom</b>	: 사각형의 아래쪽 y좌표.	<b>lwd</b>	: 선의 굵기 (테두리 및 내부 빗금)
<b>xright</b>	: 사각형의 오른쪽 x좌표.	<b>density</b>	: 내부 선들의 밀도 (내부 빗금)
<b>ytop</b>	: 사각형의 위쪽 y좌표.	<b>angle</b>	: 내부 선들의 기울기.
<b>col</b>	: 사각형의 내부 색상.		(내부 빗금 default=45)
<b>border</b>	: 사각형의 테두리 색상.	<b>main</b>	: plot의 제목, 이름.

⋮

# Visualization of CPG landscape with R

## Step 2: Gene structure plot

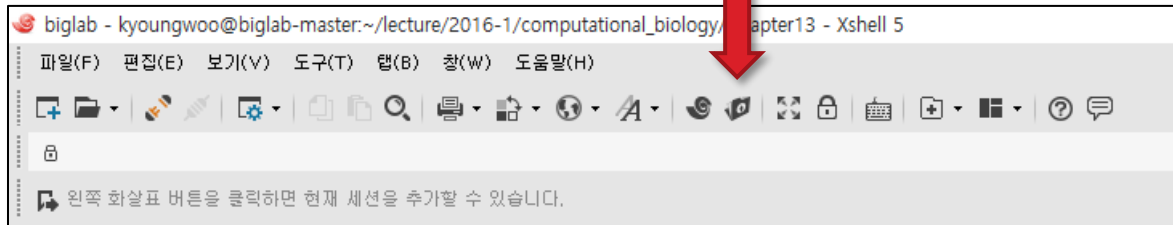
```
if (annot$category[i] == "trans") {
  # to identify the end points of transcripts
  lines(c(annot$beg[i], annot$end[i]), c(2, 2), col = "grey",
        lw = 2)
  direction <- annot$strand[i]
  if (direction == "+") {
    dir <- 2
  }
  if (direction == "-") {
    dir <- 1
  }
  # print arrows to indicate the location
  # and direction of transcript

  arrows(annot$beg[i], 1.85, annot$end[i], 1.85, col = "grey",
        code = dir, lw = 5, length = 0.1)
}

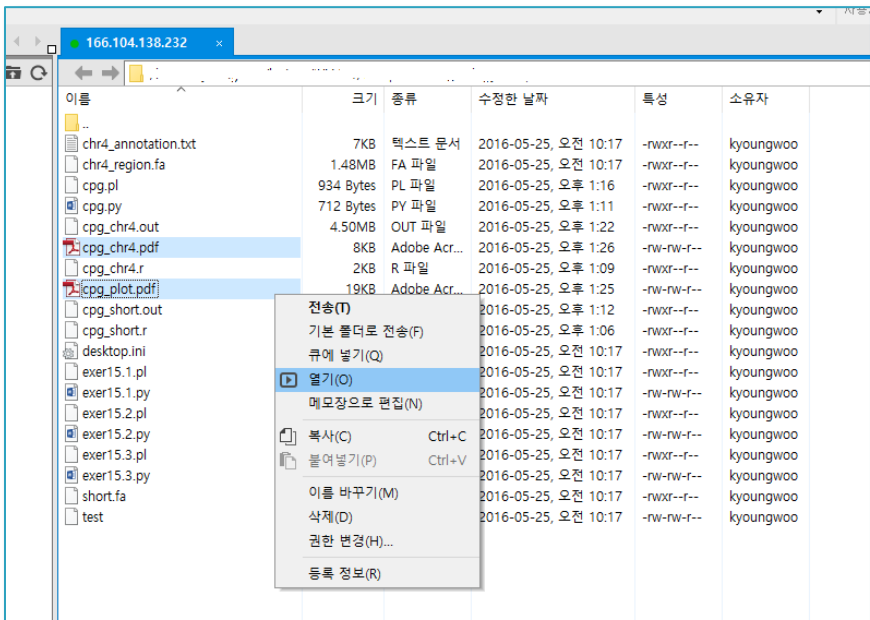
# print names of genes (this information
# can not be extracted from the
# chr4_annotation.txt file)
text(72009075 + 2e+05, 2.2, "SLC4A4")
text(72009075 + 610000, 2.2, "GC")
text(72009075 + 950000, 2.2, "NPFFR2")
text(72009075 + 1300000, 2.2, "ADAMTS3")
```

# Visualization of CPG landscape with R

```
$ Rscript cpg_chr4.r
```



# Visualization of CPG landscape with R

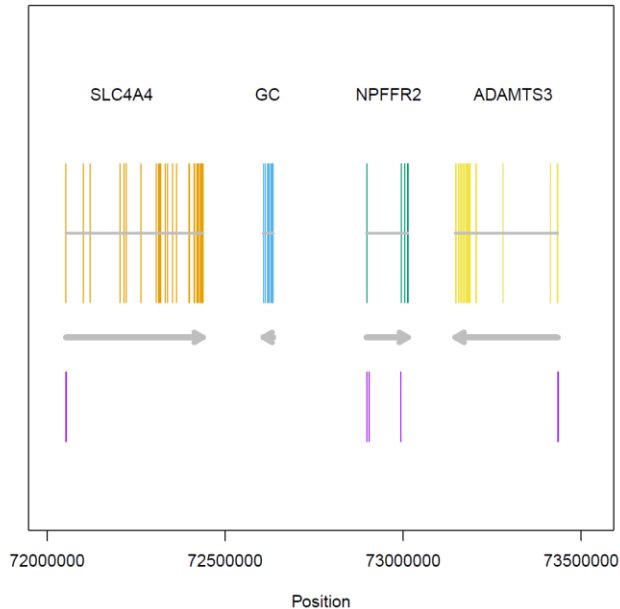




# Visualization of CPG landscape with R

cpg\_chr4.pdf

CpG island prediction



# Assignment

- ✘ In the output from Code 15.1 there is a “1” in the second column in the case that the position is part of a CpG island. Design a python script that uses the output file “cpg\_chr4.out” and prints the begin and end positions of the different CpG islands. For instance, the first CpG island that should be printed is 43070-44360
- ✘ cpg\_chr4.out의 column 2번 0 / 1 정보를 활용하여 CpG island의 시작과 끝 position을 출력하는 python 코드를 구현하세요.
- ✘ 과제 제출 기한: 11/26 Sunday 23:59 @ LMS
- ✘ 작성한 코드와 해당 코드의 결과를 캡처한 뒤 워드에 첨부(코드만 긁어와서 붙여넣지 말기), 코드에 대한 설명 간략히 작성  
워드 파일명은 n주차\_학번\_이름 형식으로 제출(e.g. 12주차\_2023123456\_김현우)